

Biosynthesis of Penicillin G from D- and L-[¹⁴C]- and [α -³H]-Valine

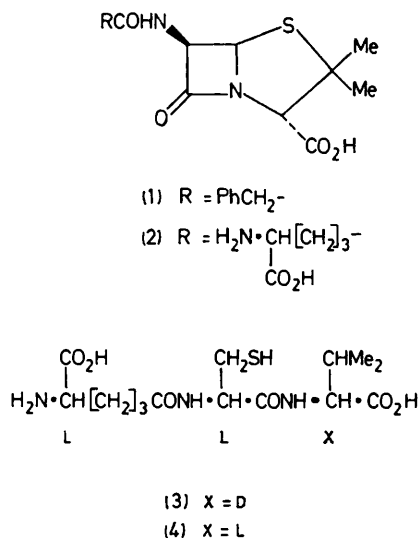
By BARRIE W. BYCROFT,* COLIN M. WELS, KENNETH CORBETT,† ADRIAN P. MALONEY,† and DAVID A. LOWE†

(Department of Chemistry, University of Nottingham, Nottingham NG7 2RD, and †Beecham Pharmaceuticals, U.K. Division, Clarendon Road, Worthing, Sussex)

Summary Excellent incorporations of both D-[1-¹⁴C]- and L-[U-¹⁴C]-valine into penicillin G by a high producing strain of *Penicillium chrysogenum* is reported; the label from D-[α -³H]-valine is not incorporated.

As an extension of our systematic investigation¹ of the biosynthesis of penicillin G (1) in a commercial high producing strain of *Penicillium chrysogenum*, we now report experiments relating to the origin of the D-valine unit.

be incorporated much less readily than the L-isomer² and there are reports that it inhibits penicillin production.³ Similarly penicillin N (2) but not cephalosporin C production is inhibited by the addition of D-valine to washed cell suspensions of *Cephalosporium* species.⁴ More recently it has been reported⁵ that (3) and not (4) is utilised by protoplasts from *Cephalosporium* for the formation of penicillin N



It is already well established that L-valine is an effective precursor of this portion of (1). However, there is conflicting evidence as to the role of D-valine which appears to

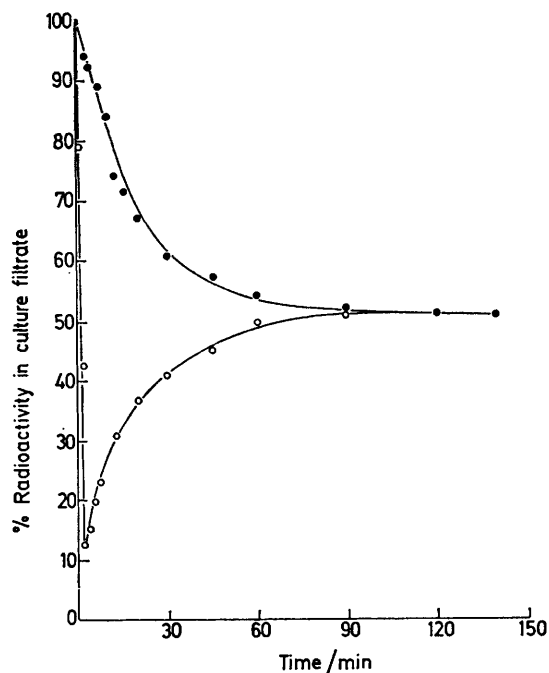


FIGURE. L-valine, ○ D-valine.

(2), suggesting that the D-centre of the valine group is generated prior to the cyclisation steps.

We have demonstrated that the uptake of ^{14}C -labelled D- and L-valine into the cells of *Penicillium chrysogenum* (Figure)† follows a characteristic pattern for each isomer. Although in all the experiments conducted the rate of uptake of D-valine was considerably slower than for the L-isomer, the final amount of radioactivity (ca. 50%) within the culture filtrate was practically the same for each isomer. In both cases a similar amount (ca. 37%) of the original activity was associated with penicillin G which was subsequently extracted and crystallised to constant activity. These experiments established that the carbon skeleton of both D- and L-valine are equally effective precursors.

In order to investigate the possibility that D-valine is incorporated directly into (1), D- $[\alpha\text{-}^3\text{H}]$ valine together with

the corresponding L-isomer was prepared by a route similar to that employed for α -tritiated cysteine.¹ Incubation of mycelium with D- $[\alpha\text{-}^3\text{H}]$ valine (3×5.3 mg; $0.21 \mu\text{Ci}/\text{mg}$) and L- $[\alpha\text{-}^3\text{H}]$ valine (3×3.0 mg; $0.40 \mu\text{Ci}/\text{mg}$) afforded (1) containing 0.20 and 0.25% activity respectively. Double labelled experiments with both isomers afforded ca. 37% incorporation of ^{14}C and again negligible incorporation of tritium.

Clearly D-valine is not incorporated directly into (1), but is possibly converted into L-valine either *via* the corresponding α -keto acid or by a mechanism not involving C-N bond cleavage. Alternatively, both D- and L-valine may proceed through a common intermediate.

(Received, 18th September 1975; Com. 1063.)

† Whole broth (ca. 300 ml) was transferred from a production fermenter during the phase of linear penicillin G production to a small scale fermenter. Glucose, vegetable oil, and sodium phenylacetate were constantly added to maintain optimum antibiotic production. The broth was equilibrated at pH 7 prior to the addition of precursor; in the case of L- $[U\text{-}^{14}\text{C}]$ valine ($0.12 \mu\text{Ci}/\text{mg}$), 3.0 mg was added after 1 h whereas with D- $[1\text{-}^{14}\text{C}]$ valine ($0.27 \mu\text{Ci}/\text{mg}$), 3.0 mg was added after an overnight incubation of ca. 18 h.

¹ B. W. Bycroft, C. M. Wels, K. Corbett, and D. A. Lowe, *J.C.S. Chem. Comm.*, 1975, 123.

² C. M. Stevens, E. Inamine, and C. W. De Long, *J. Biol. Chem.*, 1956, **219**, 405; H. R. V. Arnstein and M. E. Clubb, *Biochem. J.*, 1957, **65**, 618.

³ A. L. Demain, *Arch. Biochem. Biophys.*, 1956, **64**, 74; C. M. Stevens, P. Vohra, and C. W. De Long, *J. Biol. Chem.*, 1954, **211**, 297.

⁴ S. C. Warren, G. G. F. Newton, and E. P. Abraham, *Biochem. J.*, 1967, **103**, 902.

⁵ P. A. Fawcett, J. J. Usher, and E. P. Abraham, Proc. 2nd Internat. Symp. Genetics Industrial Micro-organisms, 1974, in the press; E. P. Abraham, personal communication.